- Zachariae H, Kragballe K, Thestrup-Pedersen K, Kissmeyer-Nielsen F. HLA antigens in methotrexate-induced liver cirrhosis. Acta Derm Venereol (Stockh) 1980; 60: 165–170.
- Nouri-Aria KT, Donaldson PT, Hegarty JE, Eddleston ALWF, Williams R. HLA A1 B8-DR3 and suppressor cell function in first degree relatives of patients with autoimmune chronic active hepatitis. J Hepatol 1985; 1:235-241.
- Jensen JR, Thestrup-Pedersen K. In vitro changes in cell-mediated immunity following tuberculin skin testing in healthy persons. Acta Pathol Microbiol Scand [C] 1983; 90: 109–116.
- Jensen JR, Thestrup-Pedersen K, From E. Fluctuations of natural killer cell activity in early syphilis. Br J Vener Dis 1983; 59:3-32.
- McFarlane IG, Wojcicka BM, Zucker GM, Eddleston ALWF, Williams R. Purification and characterization of human liver-specific membrane lipoprotein (LSP). Clin Exp Immunol 1977; 27: 381–390.
- Thestrup-Pedersen K, Ladefoged K, Andersen P. Lymphocyte transformation test with liverspecific protein and phytohaemagglutinin in patients with liver disease. Clin Exp Immunol 1976; 24: 1-6.
- Veien NK, Aldershvile J, Hardt F, Møller AM, Nielsen JO, Nyfors A, Tage-Jensen U. Immunoglobulin-bearing cells in liver biopsies from patients with psoriasis. Acta Derm Venereol (Stockh) 1978; 60: 524–526.
- Sauder DN, Bailin PL, Sundeen J, Krakauer RS. Suppressor cell function in psoriasis. Arch Dermatol 1980; 116: 51–55.

HLA-DR and DQ Antigens in Lichen planus

R. VALSECCHI,¹ M. BONTEMPELLI,² A. ROSSI,¹ P. BELLAVITA,²
A. BARCELLA,¹ A. DI LANDRO¹ and T. CAINELLI¹

¹Department of Dermatology and ²Department of Immunohematology,

Ospedali Riuniti di Bergamo, Bergamo, Italy

Valsecchi R, Bontempelli M, Rossi A, Bellavita P, Barcella A, Di Landro A, Cainelli T. HLA-DR and DQ antigens in lichen planus. Acta Derm Venereol (Stockh) 1988; 68:77–80.

A study on HLA-DR and DQ typing in 40 patients with lichen planus (26 males and 14 females) and in 92 normal blood donors of both sexes was performed. Twenty-seven patients had cutaneous lesions, 11 cutaneous and mucosal involvement and 2 patients had oral erosive lesions. Serological typing revealed a highly significant increase of HLA-DR 1 antigen in the patient group. No difference has been observed on the prevalence of DR1 antigen among the different clinical status of the disease. (Received May 8, 1987.)

R. Valsecchi, Department of Dermatology, Ospedali Riuniti, I-24100 Bergamo, Italy.

Lichen planus (LP) is a chronic papulosquamous disease that can affect skin and mucous membrane. The current literature about the pathogenesis of LP reflects a growing acceptance of a T-cell mediated immune mechanism (1–8). LP has been reported in patients with lupus erythematosus, myastenia gravis, alopecia areata and vitiligo, bullous pemphigoid and primary biliary cirrhosis. The detection of immune reactans at the basement membrane zone using direct immunofluorescence technique (9) and the recent demonstration of circulating antibodies reactive with epidermal antigens in patients with LP (10) are consistent with a proposed autoimmune pathogenesis.

Familial LP has been described (11–12) and an early HLA-study of LP patients showed HLA-A3 antigen to be significantly increased (13), but subsequent studies did not demonstrate statistically significant differences from normal controls (14–15).

Table I. Results of serological HLA-DR and DQ typing in LP patients and controls

Anti- gen	Patients (n=40)			Controls (n=92)			χ ²				
	Pres- ent	%	Ab- sent	Pres- ent	%	Ab- sent	with Yates' correction	fant degree 235-241 in K. In v. 9	Corrected p	RR	EF
DR1	26	65	14	15	16.3	77	28.64	3.35×10 ⁻⁷	3.68×10 ⁻⁶	9.53	0.58
DR2	11	27.5	29	21	22.8	71	0.12	0.72	7.92	1.28	0.06
DR3	4	10	36	21	22.8	71	2.21	0.14	1.54	0.37	-0.16
DR4	6	15	34	14	15.2	76	0.03	0.85	9.35	0.95	-0.006
DR5	8	20	32	39	42.4	53	5.16	0.02	0.22	0.33	-0.38
DR6	4	10	36	3	4.3	89	1.36	0.24	2.64	3.29	0.06
DR7	10	25	30	20	26.0	72	0.034	0.85	9.35	1.20	0.04
DR8	1	2.5	39	7	7.6	85	0.54	0.46	5.06	0.31	-0.05
DQ1	32	80	8	57	61.9	35	3.35	0.07	0.77	2.45	0.47
DQ2	13	32.5	27	41	44.6	51	1.22	0.27	2.97	0.59	-0.21
DQ3	15	37.5	25	46	50	46	1.28	0.26	2.86	0.60	-0.25

Because it is known that DR antigens can control the susceptibility and the expression of some immunologic disorders, we performed HLA-DR typing on patients with LP.

DQ antigens, which are class II antigens closely related to DR, were also typed during the study.

MATERIALS AND METHODS

Forty LP patients seen in our department, from October 1985 to October 1986, had HLA-DR and DQ typing carried out. There were 14 females and 26 males ranging in age from 30 to 60 years. Twenty-seven patients had cutaneous lesions, 11 cutaneous and mucosal involvement and 2 patients had oral erosive LP. No patient was suspected of having drug-induced LP. Ninety-two normal blood donors of both sexes were studied during the same time period, using the same methods and antisera. None had a personal and family history of LP. All of the subjects lived in the same geographic area as the patients and HLA testing was carried out without knowledge of the clinical status of the patients. HLA-DR and DQ typing was performed by the microlymphocytotoxicity test of Terasaki & McClelland for the following specificities: DR1, DR2, DR3, DR4, DR5, DR6, DR7, DQ1, DQ2, DQ3.

The significance of possible deviations between the frequencies observed in the patients and in the control subjects has been evaluated by means of χ^2 analysis with the correction of Yates for discontinuities. p values were then corrected by the number of antigens tested. The value of the strength and rank of association was obtained by calculating the relative risk (RR). We have also evaluated the etiological fraction (EF); EF value indicates how much of a disease is "due to" the disease-associated factor. It is computed from RR value and the frequency of the antigen in patients.

RESULTS

The results of HLA-DR and DQ typing are summarized in Table I, together with the calculated χ^2 , p, corrected p values, RR and EF. HLA-DR1 was found to be significantly increased in LP patients with p and corrected p values of 3.35×10^{-7} and 3.68×10^{-6} respectively. The RR for patients with DR1 of developing LP was 9.53 and the EF value was 0.58. No difference has been observed on the prevalence of DR1 antigen among the different LP clinical status.

DISCUSSION

Our study on HLA-DR and DQ typing shows a striking statistically significant increase in the frequency of DR1 antigen in LP patients. These data are in partial agreement with a concomitant study (16), where the serological typing for HLA Class II antigens in LP revealed a highly significant increase of HLA-DR1 and MT1 (DQ1) antigens. In our study, in fact, we have obtained furthermore a significantly high frequency of HLA-DR1. The frequency of DQ1 antigen is higher in the group of patients. The difference, however, with respect to the control subjects is not statistically significant, but near to the significance level.

Our results really are apparently conflicting with the Mayo Clinic report, and this fact may be due, at least in part, to the different number of patients studied.

The frequency of HLA-DQ1 antigen in patients and in controls is similar in the two studies, showing that the two populations are genetically comparable. Moreover, the linkage disequilibrium between DR1 and DQ1 antigens is very frequent, but not absolute.

On the other hand our and Powell's results (16) support the hypothesis that LP is probably linked more with DR1 antigen rather than with DQ1. We also must remember that the locus-DR is frequently associated with most of HLA-linked immune diseases.

Previous HLA studies of patients with LP have shown an increased frequency of HLA-A3. This antigen, controlled by the correspondent gene of Locus A, is known to be in linkage disequilibrium with DR1 (17). So we can suggest that an increased frequency of A3 may reflect a stronger association with a Class II antigen.

During the last two years DR1 antigen has been linked with reduced serological response to Epstein-Barr viral antigen (18) and MT3 (DRW53) has been shown to be involved in the presentation of mumps viral antigen to T-cell (19). These last data suggest that different Class II antigens may be involved in the afferent limb of the immune response to alloantigens.

Ir genes are thought to be associated to the DR locus and are expressed through Ia antigens on different cells (endothelial cells, macrophages, T-lymphocytes and Langerhans' cells of the epidermis). Some of these Ia-like antigen-carrying cells are involved in LP. Variations in the relative number of T-cell subsets with the age of LP lesions have been described (2, 3, 5), thus suggesting an antigen specific education of inducer T-cells in early LP lesions, whereas in late lesions cytotoxic T-cell could destroy keratinocytes (3, 5, 7, 8).

Our study has shown that in LP patients there is a statistically significant increase of HLA-DR1 antigen; this result could support the hypothesis that, in some predisposed subjects, epithelial Langerhans' cells may process a foreign antigen (virus, drug, allogeneic cells, chemical substances) in a particellar way and induce a local immune response.

The antigen activated lymphocytes in the dermis may secrete the recombinant human gamma interferon, which causes some changes: expression of HLA-DR by keratinocytes and an increased rate of differentiation of these cells with formation of a thickened granular cell layer.

Moreover, the observed destruction of basal cell layer in LP could be explained in terms of keratinocytes expressing a foreign HLA-DR on their surface as in graft-versus-host disease, or presenting an alloantigen with autologous HLA-DR. As a consequence the keratinocytes are recognized as foreign by the immunocompetent cells and become targets for cytotoxic T-cells.

REFERENCES

- Braathen LR, Dahl BL, Mellbye OJ. Immunological in situ identification of the lymphoid cells in the skin infiltrate of lichen planus. Clin Exp Dermatol 1979; 4: 175–181.
- Bhan AK, Harrist TJ, Murphy GF, Mihm MLJ. T-cell subsets and Langerhans cells in lichen planus: in situ characterization using monoclonal antibodies. Br J Dermatol 1981; 105:617-622.
- 3. De Panfilis G, Manara GC, Ferrari C, Manfredi G, Allegra F. Imbalance in phenotypic expression

- of T-cell subpopulations during different evolutional stages of lichen planus. Acta Derm Venereol (Stockh) 1983; 63: 369-373.
- De Panfilis G, Manara GC, Sansoni P, Allegra. F. T-cell infiltrate in lichen planus. Demonstration of activated lymphocytes using monoclonal antibodies. J Cutan Pathol 1983; 10: 52-58.
- De Panfilis G, Manara GC, Allegra F. Remarks on early versus late lichen planus. Arch Dermatol Res 1981; 270: 163-166.
- Gianotti B, de Panfilis G, Manara GC, Allegra F. Macrophage-T-lymphocyte interaction in lichen planus. An electron microscopic and immunocytochemical study. Arch Dermatol Res 1983; 275: 35-40.
- 7. Gones MA, Schmitt DS, Souteyrand, P, Ohrt C, Brochier J, Thivolet J. Lichen planus and chronic graft-versus-host reaction. In situ identification of immunocompetent cell phenotypes. J Cutan Pathol 1982; 9: 249-258.
- 8. Ragaz A, Ackerman AB. Evolution, maturation and regression of lesions of lichen planus. Am J Dermatopathol 1981; 3:5-25.
- 9. Abell E, Presbury DGL, Marks R, Ramnarain D. The diagnostic significance of immunoglobulin and fibrin deposition in lichen planus. Br J Dermatol 1975; 1:17-24.
- Olsen RG, Du Plessis DP, Joy Schulz E, Camisa C. Indirect immunofluorescence microscopy of lichen planus. Br J Dermatol 1984; 110: 9-15.
- Kofoed ML, Wantzin GL. Familial lichen planus. J Am Acad Dermatol 1985; 13:50-54.
- 12. Mahood JM. Familial lichen planus. A report of nine cases from four families with a brief review of the literature. Arch Dermatol 1983; 119: 292-294.
- Lowe NJ, Cud Worth AG, Woodrow JC. HLA antigens in lichen planus. Br J Dermatol 1976; 95: 169-171.
- 14. Saurat JH, Lemarchand F, Hors J, Numes-Roldan A, Gluckman E, Dausset J. HLA markers and lymphocytotoxins in lichen planus. Arch Dermatol 1977; 113: 1719-1724.
- 15. Veien NK, Risum R, Jorgensen P, Svejgaard A. HLA antigens in patients with lichen planus. Acta Derm Venereol (Stockh) 1979; 59: 205-209.
- 16. Powell FC, Rogers RS, Dickson ER, Breanndan Moore S. An association between HLA-DR1 and lichen planus. Br J Dermatol 1986; 114: 473-478.
- 17. Batchelor JR. Joint report on DR1 in histocompatibility testing (ed. by Terasaki, p. I), p. 508. Los Angeles, 1980, February 4-10.
- 18. Cohen JHN, Vischer TL, Carquin J, Blanchard F, von Fliedner VE, Jeanett M. HLA-DR antigens and the antibody response against Epstein-Barr virus. Tissue Antigens 1984; 23: 156-159.
- 19. Ball EJ, Stastny P. Antigen specific HLA-restricted human T-cell lines. I. An MT-like restriction determinant distinct from HLA-DR. Immunogenetics 1984; 19:13-19.

Ultrastructure of Angiokeratoma Vulvae

T. KARLSMARK, 1 K. WEISMANN 1 and T. KOBAYASI2

Department of Dermato-venereology, Bispebjerg Hospital, University of Copenhagen, and

²Department of Dermatology, Rigshospital, University of Copenhagen, Denmark

Karlsmark T, Weismann K, Kobayasi T. Ultrastructure of angiokeratoma vulvae. Acta Derm Venereol (Stockh) 1988; 68: 80-82.

A case of vulvar angiokeratoma studied by electron microscopy is described. The patient, a 51-year-old woman, had noticed eruptions for the last 15 years, though without symptoms. Osmiophilic bodies with myelin-like figures were found in the vascular endothelial cells. The findings support previous opinion that vulvar angiokeratoma is a variant of scrotal angiokeratoma. (Received July 14, 1987.)

T. Karlsmark, Department of Dermatology, Bispebjerg Hospital, Bispebjerg Bakke 23, DK-2400 Copenhagen, Denmark.

Angiokeratoma vulvae is probably a variant of scrotal angiokeratoma Fordyce (1). Few papers have described the condition, though it seems to be more common than generally