Dithranol (Anthralin) and 10-Butyryl Dithranol (Butantrone) Do Not Morphologically Transform Cultured C3H 10T1/2 C18 Mouse Embryo Fibroblasts

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The ability of dithranol and 10-butyryl dithranol to induce morphological cell transformation was studied in cultured C3H 10T1/2 C18 fibroblasts. The cells were incubated with different concentrations of the test compounds for 48 h and cultured for 5 weeks thereafter. At the end of the culture period the cultures were fixed, stained and examined for the presence of transformed foci. Dithranol and 10-butyryl dithranol did not increase the formation of transformed foci, while the positive control compound, 7,12-dimethylbenz(a)anthracene (DMBA), induced a high frequency of transformations significantly different from controls. Thus the in vitro cell transformation model with uninitiated C3H 10T1/2 C18 fibroblasts is not able to detect the weak tumorigenic action of dithranol and 10-butyryl dithranol which has been observed in mouse skin. Key words: Cell transformation; Carcinogenesis; In vitro.

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Dithranol is a widely used and effective antipsoriatic drug, which has not been shown to cause skin cancer in psoriasis patients (1–3). However, it is a well-known tumor promoter in in vivo experimental models, like mouse skin (4-9), and it has also been shown to act as a cocarcinogen when administered simultaneously with benzo(a)pyrene (BaP) (7). 10-butyryl dithranol (butantrone), an acutely less irritating derivative of dithranol, has a lower tumor-promoting activity than dithranol in mouse skin (8, 9). Both compounds have also shown a weak direct tumorigenic activity by inducing squamous cell papillomas even without initiation with 7,12-dimethylbenz(a)anthracene (DMBA) (4-6, 8, 9). In addition, there is evidence that dithranol and, to a lesser extent, 10-butyryl dithranol are able to interact with DNA. Dithranol was reported to induce DNA strand breaks in human leucocytes in vitro (10). Dithranol and 10-butyryl dithranol were mutagenic in Salmonella typhimurium TA1537 both with and without metabolic activation. They also showed a clastogenic activity by increasing the number of chromosomal gaps and breaks in human lymphocytes in vitro (11).

The purpose of this study was to examine if dithranol and butantrone show a direct transforming potential in cultured C3H 10T1/2 C18 mouse embryo fibroblasts. This test system allows the expression of transformed foci of dense multilayered cells on a non-transformed contact-inhibited monolayer.

MATERIALS AND METHODS

Test compounds

Dithranol (mol. wt 226.2) was obtained from Bayer (Leverkusen, Germany) and purified by recrystallizing from a mixture of acetic acid and water (90:10). 10-butyryl dithranol (mol. wt 296.3) was synthesized according to Mustakallio et al. (12) and recrystallized from a mixture of isopropanol and acetonitrile. The test substances were dissolved in dimethylsulphoxide (DMSO) immediately prior to the treatment.

Cell culture

C3H 10T1/2 C18 cells at the passage number of 11–15 were grown in Dulbecco's modified essential medium with Earle's salts (DMEM) containing 5% fetal calf serum (Gibco, Grand Island, NY, USA) and gentamycin (25 mg/ml). Cells were maintained at exponential growth conditions in a humidified incubator containing 10% CO₂ in air at 37°C.

Cytotoxicity and cell transformation assays

The concentrations for the transformation assays were selected in preliminary cytotoxicity tests, where cell survival and colony forming efficiency were measured. In the final transformation assays, a solvent control (0.1% DMSO), five concentrations (0.05–0.5 $\mu g/ml$) of the test compounds and a positive control (DMBA, 0.05 $\mu g/ml$) were used. The final concentration of DMSO was 0.1%. The cells were plated on 60 mm Petri dishes (2000 cells/dish, 15–30 replicates), exposed for 48 h and cultured for 5 weeks. The growth medium was changed twice weekly. At the end of the culture period the cultures were fixed with methanol, stained with 5% aqueous Giemsa and scored for the presence of morphologically transformed type II and type III foci ($\varnothing > 2$ mm) according to Reznikoff (13). Statistical significance of the treatments was calculated using Fisher exact test.

RESULTS

Cytotoxicity of dithranol and 10-butyryl dithranol, expressed as colony-forming efficiency, is shown in Fig. 1. Dithranol was slightly more toxic to C3H 10T1/2 C18 cells than 10-butyryl dithranol.

The numbers of transformed foci observed after the culture period of 5 weeks are presented in Table I. Dithranol and 10-butyryl dithranol did not significantly or dose-dependently increase the formation of transformed foci, while the positive control DMBA induced a high frequency of transformations, significantly different from controls.

DISCUSSION

Both the therapeutic and toxic effects of dithranol are related to the formation of free radicals during dithranol autoxidation (14–18). In addition to the highly reactive dithranol-free radical (10-anthranyl radical), reactive oxygen species (singlet oxygen, superoxide anion radical, hydroxyl radical) are the biologically most significant radical species formed during dithranol

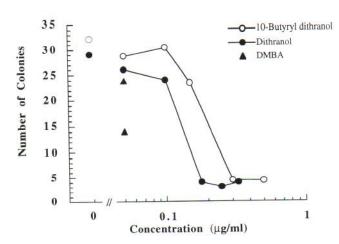


Fig. 1. Cytotoxicity of dithranol and 10-butyryl dithranol to C3H 10T1/2 C18 cells. Two hundred cells were plated on 60 mm Petri dishes, treated with test compounds for 48 h, and incubated thereafter for 7 days. Mean numbers of colonies (containing at least 50 cells) per dish, 5 replicates.

autoxidation. The ability of dithranol and 10-butyryl dithranol to react with DNA (10, 11) is obviously related to the formation of the free radicals. However, the compounds were not able to transform uninitiated C3H 10T1/2 C18 cells in this study.

Dithranol has not uniformly shown genotoxic activity in different models for genotoxicity. Dithranol was unable to induce chromosomal mutations in yeast (Saccharomyces cerevisiae) or ascomycete (Ophistoma multiannulatum) (19). Similarly, it did not induce forward mutagenesis, sister chromatide exhanges or structural chromosomal changes in V79 Chinese hamster

Table I. The effect of dithranol and 10-butyryl dithranol on the induction of transformed foci in C3H 10T1/2 C18 cells

The cells were treated with the test compounds for 48 h and cultured for 5 weeks thereafter (15–30 dishes per concentration).

Treatment	Concentration		Number of transformed foci			
	μg/ml	(μΜ)	Туре			Total
			П	III	II+III	foci/dish
Control 1	0	(0)	7	0	7	0.30
Dithranol	0.05	(0.221)	5	1	6	0.25
	0.10	(0.442)	5 5	0	5	0.26
	0.18	(0.796)	3	0	3	0.20
	0.25	(1.105)	16	0	16	0.89
	0.33	(1.459)	5	0	5	0.33
DMBA 1	0.05	(0.195)	19	21	40	2.22*
Control 2	0	(0)	7	0	7	
10-butyryl dithranol	0.05	(0.169)	5	0	5	0.17
	0.10	(0.337)	9	0	9	0.36
	0.15	(0.506)	12	0	12	0.43
	0.30	(1.012)	5	0	0	0.24
	0.50	(1.687)	4	1	5	0.29
DMBA 2	0.05	(0.195)	24	42	66	3.30*

Statistics: p < 0.0005, Fisher exact probability test.

lung fibroblasts (20). In some cases the use of too high (toxic) concentrations has confused the interpretation of the results (21, 22).

In previous cell transformation studies dithranol has been shown to be active when initiated cells are used (in vitro substitutes of tumor promotion studies). Dithranol transformed virally infected Swiss 3T3 mouse fibroblasts to form colonies in soft agar (23). Baturay & Trombetta (24) showed that dithranol transformed Balb/c-3T3 cells after initiation with either BaP, which needs metabolic activation for its carcinogenic action, or β-propiolactone (BPL), a direct-acting carcinogen. Moreover, when the cells were treated with one of the initiators and dithranol at the same time, the cells were transformed with BaP, but not with BPL. In this study, however, without initiation or simultaneous treatment with initiators, no cell transformation was observed. Therefore, the in vitro cell transformation model with uninitiated C3H 10T1/2 C18 cells does not predict or is not sensitive enough to detect the observed weak direct tumorigenic action of dithranol and 10-butyryl dithranol in mouse skin.

REFERENCES

- Swanbeck G, Hillström L. Analysis of etiological factors of squamous cell skin cancer of different locations. Acta Derm Venereol (Stockh) 1971; 51: 151–156.
- Bridges BA, Greaves M, Polani PE, Wald N. Do treatments available for psoriasis patients carry a genetic or carcinogenic risk? Mutation Res 1981; 86: 279–304.
- Stern RS, Lange R. Cardiovascular disease, cancer and cause of death in patients with psoriasis: 10 years prospective experience in a cohort of 1380 patients. J Invest Dermatol 1988; 91: 197–201.
- Bock FG, Burns R. Tumor-promoting properties of anthralin (1,8,9anthratriol). J Natl Cancer Inst 1963; 30: 393–397.
- Yasuhira K. Skin papillomas produced by anthralin painting after urethane initiation in mice. Gann 1968; 59: 187–193.
- Van Duuren BL, Goldschmidt BM. Cocarcinogenic and tumorpromoting agents in tobacco carcinogenesis. J Natl Cancer Inst 1976; 56: 1237–1242.
- Van Duuren BL, Segal A, Tseng S-S, Rusch GM, Loewengart G, Maté U, et al. Structure and tumor-promoting activity of analogues of anthralin (1,8-dihydroxy-9-anthrone). J Med Chem 1978; 21: 26–31.
- Männistö PT, Vaissi L, Mustakallio KK, Viluksela M, Kosma V-M, Collan Y. Tumor-producing activity of dithranol (anthralin) and two of its 10-acyl analogs in the dorsal skin of female NMRI mice. J Pharmacol Exp Ther 1984; 229: 255–260.
- Viluksela M, Puotunen E, Newman AJ, Männistö PT. Tumorproducing and skin irritating activity of dithranol (anthralin) and its 10-acyl analogues in SENCAR mice. Carcinogenesis 1986; 7: 1755–1760.
- Birnboim HC. DNA strand breaks in human leukocytes induced by superoxide anion, hydrogen peroxide and tumor promoters are repaired slowly compared to breaks induced by ionizing radiation. Carcinogenesis 1986; 7: 1511–1517.
- Männistö PT, Kirkland D, Viluksela M, Tikkanen L. Toxicological studies with dithranol and its 10-acyl analogues. Arch Toxicol 1986; 59: 180–185.
- Mustakallio KK, Pippuri AK, Honkanen E. Dihydroxyacylanthrons having anti-psoriatic activity. US Patent 1981; 4.299.846.
- Reznikoff CA. Quantitative and qualitative studies of chemical transformation of cloned C3H mouse embryo cells sensitive to post-confluence inhibition of cell division. Cancer Res 1973; 33: 3239–3249.
- Martinmaa J, Vanhala L, Mustakallio KK. Free radical intermediates produced by autoxidation of 1,8-dihydroxy-9-anthrone (dithranol) in pyridine. Experientia 1978; 34: 872.

- Finnen MJ, Shuster S, Lawrence CM. Inhibition of dithranol inflammation by free-radical scavengers. Lancet 1984; ii: 1129– 1130
- Shroot B, Brown C. Free radicals in skin exposed to dithranol and its derivatives. Arzneim-Forsch 1986; 36: 1253–1255.
- Müller K, Kappus H. Hydroxyl radical formation by dithranol. Biochem Pharmacol 1988; 37: 4277–4280.
- Fuchs J, Packer L. Investigations of anthralin free radicals in model systems and in skin of hairless mice. J Invest Dermatol 1989, 92: 677–682.
- Zetterberg G, Swanbeck G. Studies on dithranol and dithranol-like compounds. II. Mutagenicity. Acta Derm Venereol (Stockh) 1971; 51: 45–49.
- Kinsella AR. Elimination of metabolic co-operation and the induction of sister chromatid exchanges are not properties common to

- all promoting or co-carcinogenic agents. Carcinogenesis 1982; 3: 499–503.
- McCann J, Choi E, Yamasaki E, Ames BN. Detection of carcinogens as mutagens in the Salmonella/microsome test: assay of 300 chemicals. Proc Natl Acad Sci USA 1975; 72: 5135–5139.
- Bernd A, Holzmann H, Marsch WC, Kurelec B, Britvic S, Müller WEG. Antimutagenic potency of the cytotoxic and anti-psoriatic compound anthralin (Cignolin). Pharmacol Res Commun 1987; 19: 367–378.
- Daya-Grosjean L, Sarasin A, Monier R. Effect of tumor promoters on soft-agar growth of Swiss 3T3 cells infected with SV40 tsA mutants. Carcinogenesis 1982; 3: 833–835.
- Baturay NZ, Trombetta LD. Cocarcinogenic and tumor-promoting capabilities of anthralin. Arch Dermatol Res 1988; 280: 443–450.