

Appendix S1

MATERIALS AND METHODS

After obtaining informed consent, ethylenediaminetetraacetic acid (EDTA)-blood samples, and skin biopsies were taken from the index case. The study was approved by the ethics committee of the University of Freiburg, and conducted according to the principles of the Declaration of Helsinki.

Mutational analysis of the coding region and exon-intron boundaries of the *DST* gene was performed as described previously (9). Sanger sequences for the DNA were compared with reference sequences from the NCBI Entrez Nucleotide database

(NM_001723.4; NG_029322.2). The primers used in this study are listed in **STable I**.

Normal skin specimens, obtained with the informed written consent of individuals who underwent surgery, were used for cell isolation and immunostaining. Keratinocytes derived from normal and patient's skin were immortalized with the HPV E6 and E7 genes (plasmids were gifts from Fernando Larcher and Stephanie Löffek) and cultured in keratinocyte growth medium (Invitrogen, Karlsruhe, Germany).

Immunofluorescence staining of the skin and cells was performed as described previously (10, 11). Confluent cell monolayers were lysed and immunoblotted as described previously (12). Primary and secondary antibodies are listed in **STable II**.

STable I. Primers used in this study

Primer name	Sequence 5'-3'	Primer name	Sequence 5'-3'	Primer name	Sequence 5'-3'
<i>Genomic DNA</i>		BPAG1e-ex19R	ggaagatgaggagcatgactg	<i>cDNA</i>	
BPAG1e-ex1F	tgccacttttcaccgtaga	BPAG1e-ex20F	agtgcacaaaggctaaggagc	GAPDH	gaaggtgaaggtcggagtc
BPAG1e-ex1R	aagtgctgcccccaaaaag	BPAG1e-ex20R	aaacactgagtgaaatgacttg		ttgaggtcaatgaaggggtc
BPAG1e-ex2F	gcagcactttcaggaatgg	BPAG1e-ex21F	gtaatgactgtgccacgac	DST	ttgacaccccttagatagtaagaac
BPAG1e-ex2R	caaacgcacagtgactcctc	BPAG1e-ex21R	ccctcccccttttctctca		tcacccctgactgaaatcgc
BPAG1e-ex3F	tttcagggtgtttgaaacttg	BPAG1e-ex22F	gctgtctgaaggcctagttaatg	Col 17A	tctgaaggggatcatcaag
BPAG1e-ex3R	ttcaaatgccatgaaacag	BPAG1e-ex22R	catgttttgcctcccaaac		accctgaactcggataggt
BPAG1e-ex4F	ttcactctgtttacaagtgcc	BPAG1e-ex23aF	ttggtgatagcgctgttctg	ITGA6	ggcgggttatgtcctgagt
BPAG1e-ex4R	gggtccagttgatccttcagacag	BPAG1e-ex23aR	tgcgaaaattcaggagggttc		catgtctcagctctccacca
BPAG1e-ex5/6F	attgggtgatgagatgcag	BPAG1e-ex23bF	taccgcagggaaactgaaac	ITGB4	gaggtaggtccaggacggg
BPAG1e-ex5/6R	tcttgattctggatgtattcttag	BPAG1e-ex23bR	gcctttctattggcagctgtt		ctgatcaaggctccaggag
BPAG1e-ex7/8F	tctgtttccctatctatattggc	BPAG1e-ex23cF	gaaaatgcattgccagtgtg	FERMT1	cattactgatccctaaacttg
BPAG1e-ex7/8R	caactctcagcaataaagaatctg	BPAG1e-ex23cR	catggcccttgattctg		ttggatttgtctttttgc
BPAG1e-ex9F	cgatatcaaaaggtatgtgtgcc	BPAG1e-ex23dF	gaagaagccatgcaagaagc	FERMT2	gaggggcctctatcactcc
BPAG1e-ex9R	tcctgcaagttagccaaatc	BPAG1e-ex23dR	tgtgtcgtgtagctgactt		tctgctgtgttgtagacag
BPAG1e-ex10F	ggggtcacataggaccttg	BPAG1e-ex23eF	gcactgaaaattcaggcaga	ITGA3	ctgggtgctgtactctgtgc
BPAG1e-ex10R	gccataaaaatacaaaagttctgctc	BPAG1e-ex23eR	ttgctgcatttgttcacgat		gctgtctctgacccctgac
BPAG1e-ex11/12F	aaagtgtcaagggcatggag	BPAG1e-ex23fF	cagcagaggagttcgggaag	ITGB1	taagatcaggggagccacag
BPAG1e-ex11/12R	ccctcaaccttcagagggc	BPAG1e-ex23fR	ctggcactcctcactgtca		tcagaattgattgtgctca
BPAG1e-ex13F	tgcaagtgttttcagaggg	BPAG1e-ex23gF	aaagagcacagccaaagactg		
BPAG1e-ex13R	caaaactctcagcacatatgaaaagg	BPAG1e-ex23gR	caatgagccaatcacattcaa		
BPAG1e-ex14F	ttggcatataccacacgacc	BPAG1e-ex24aF	ctgaacctccttggtttga		
BPAG1e-ex14R	cctaagattgagatcccattg	BPAG1e-ex24aR	attgaggtggcttctgctag		
BPAG1e-ex15F	tgcttgactagctgtgtccc	BPAG1e-ex24bF	aggcagttgggtggaagctaa		
BPAG1e-ex15R	tgtgcaaaaactgacattaagaag	BPAG1e-ex24bR	tgctggatggctcatgtaaa		
BPAG1e-ex16F	cattgtgtcttaatgtgaaatgtg	BPAG1e-ex24cF	ttgtgagagcattcgtgttc		
BPAG1e-ex16R	gccagccaataatgatattc	BPAG1e-ex24cR	actaacggcctcagcaaga		
BPAG1e-ex17F	tcagtgttccaaggtgaagaac	BPAG1e-ex24dF	caggaaggcctcatcact		
BPAG1e-ex17R	gagatcacagagacacaaatgg	BPAG1e-ex24dR	gttttggcaatgaggaca		
BPAG1e-ex18F	gcaaaatttataggctgaacg	BPAG1e-ex24eF	aaatgggaatccgatgtttg		
BPAG1e-ex18R	tcctccaagtaagaagaataggg	BPAG1e-ex24eR	tttgaaggcattaaatctatgTaa		
BPAG1e-ex19F	catctccacctcctgatg				

Total RNA was isolated from keratinocytes using RNAeasy® FFPE kit (QIAGEN, Hilden, Germany), transcribed into cDNA (Fermentas, St Leon-Rot, Germany), and subjected to quantitative

real time PCR (qPCR) using iQTM SYBR® Green Supermix, Biorad CFX96 and BioRad CFX Manager Software (version 1.5). Primers are listed in STable II.

STable II. Antibodies used in this study

Primary antibodies/Antigen	Clone/Cat. no.	Company	Application and dilution
BPAG1	279	Cosmo BIO	IF 1:50
Phalloidin, TRITC-conjugated	FAK100	Millipore (Darmstadt, Germany)	IF 1:1000
Collagen XVII	NC16A, polyclonal rabbit	Schäcke et.al. (S1)	IF 1:1000 IB 1:1000
Collagen XVII	NC16A-3	Abcam (Cambridge, UK)	IF 1:1000
Collagen XVII	Endo 2	Franzike et al. (S2)	IB 1:1000
Plectin	31	BD Biosciences	IF 1:250
GAPDH	6C5	Merck (Darmstadt, Germany)	IB 1:2000
Integrin α6	GOH3	Progen (Heidelberg, Germany)	IF 1:50
Integrin β4	3E1	Millipore (Darmstadt, Germany)	IF 1:100
Integrin α3	P1B5	Chemicon	IF 1:100
Integrin β1	4B7R	Abcam (Cambridge, UK)	IF 1:50
Keratin 5/6	D5/16 B4	Dako (Hamburg, Germany)	IB, IF 1:500
Keratin 5	Polyclonal, ab53121	Abcam (Cambridge, UK)	IF 1:1000
Keratin 14	LL002	Abcam (Cambridge, UK)	IB 1:500 IF 1:100
Keratin 15	EPR1614Y	Abcam (Cambridge, UK)	IB 1:400 IF 1:100
Fibronectin	Polyclonal	Abcam (Cambridge, UK)	IB 1:2000 IF 1:1000
Laminin	Polyclonal	Abcam (Cambridge, UK)	IF 1:1000
Secondary antibodies		Company	Dilution
Alexa Fluor® 488 goat anti-mouse IgG		Invitrogen, Darmstadt, Germany	1:1000
Alexa Fluor® 488 goat anti-rabbit IgG		Invitrogen, Darmstadt, Germany	1:1000
Alexa Fluor® 594 goat anti-rabbit IgG		Invitrogen, Darmstadt, Germany	1:1000
Alexa Fluor® 568 goat anti-mouse IgG		Invitrogen, Darmstadt, Germany	1:1000
Horseradish peroxidase labeled goat anti-mouse		Merck, Darmstadt, Germany	1: 5000
Horseradish peroxidase labeled goat anti-rabbit		KPL, Gaithersburg, MA, USA	1:10000

IB: immunoblotting; IF: immunofluorescence staining; Cat. no.: catalogue number.

SUPPLEMENTARY REFERENCES

- S1. Schäcke H, Schumann H, Hammami-Hausli N, Raghunath M, Bruckner-Tuderman L. Two forms of collagen XVII in keratinocytes. A full-length transmembrane protein and a soluble ectodomain. *J Biol Chem* 1998; 273: 25937–25943.
- S2. Franzke C-W, Tasanen K, Schäcke H, Zhou Z, Tryggvason K, Mauch C, et al. Transmembrane collagen XVII, an epithelial adhesion protein, is shed from the cell surface by ADAMs. *EMBO J* 2002; 21: 5026–5035.