The in vitro activity of a new triazole R126638 against Malassezia yeasts was lower with that of ketoconazole. With the agar dilution technique, minimal inhibitory concentrations were lower for R126638 compared with ketoconazole against Malassezia globosa, M. obtusa, M. slooffiae, M. restricta and two strains of M. sympodialis. On human stratum corneum in vitro, both R126638 and ketoconazole were very effective in reducing the production of hyphae from 15% to 1% with R126638 and to 2% with ketoconazole. Scanning electron microscopy did not reveal obvious surface differences between untreated cultures and cultures exposed to ketoconazole or R126638 in the concentration range 0.01–1 µg/ml. However, transmission electron microscopy showed partial to complete necrosis of the cytoplasmic organelles of Malassezia yeasts. The combined scanning electron microscopy and transmission electron microscopy findings confirm earlier observations of the “mummifying” effect of azoles against Malassezia spp. In conclusion, R126638 is an interesting new triazole with high activity against the Malassezia yeasts, which are involved in pityriasis versicolor and seborrhoeic dermatitis. Key words: Malassezia species; R126638; ketoconazole; MIC; hyphae formation.

(Accepted January 4, 2006.)


Jan Faergemann, MD, PhD, Department of Dermatology, Sahlgrenska University Hospital, SE-413 45 Göteborg, Sweden. E-mail: Jan.Faergemann@derm.gu.se

R126638 is a broad-spectrum antifungal belonging to the class of triazoles (1, 2). It has excellent potential for oral and topical treatment of fungal infections of the skin, hair, nails, oral and genital mucosa. In vitro data has demonstrated R126638 activity against dermatophytes (Trichophyton spp., Microsporum canis, Epidermophyton floccosum), yeasts (Candida spp. and Malassezia spp.) and many other fungi. Furthermore, animal experiments have provided evidence for a potent therapeutic effect of R126638 that has proved to be 4- to 8-fold superior to that of itraconazole, especially for superficial fungal infections (1). The strong activity of R126638 is ascribed to its prominent affinity for fungal cytochrome P450, which is involved in the biosynthesis of ergosterol from lanosterol. Ergosterol is a vital cell membrane component in fungi, and its inhibition by R126638 leads to abated fungal growth (3).

Activity of R126638 against several dermatophytes and M. globosa was demonstrated in the stratum corneum of healthy volunteers taking 100 mg or 200 mg R126638 daily for one week (4).

Ketoconazole is a broad-spectrum, orally and topically active imidazole derivative (5). It has a high activity against Malassezia in vitro and is also effective in the treatment of various Malassezia-associated diseases.

The genus Malassezia can be divided into seven species (6). M. furfur, M. sympodialis, M. globosa, M. obtusa, M. restricta and M. slooffiae are the lipophilic members of the genus. M. pachydermatis is the non-lipophilic member of the genus and isolated primarily from animals.

The lipophilic Malassezia species are members of the normal skin flora (7, 8). However, they are not only harmless saprophytes but also opportunistic pathogens. They are the aetiological agent or play an important role in pityriasis versicolor, Pityrosporum (Malassezia) folliculitis, seborrhoeic dermatitis and atopic dermatitis (7, 9, 10). Even systemic infections with these yeasts have been reported (8).

In a paper by Gupta et al. (11), the in vitro susceptibility (minimal inhibitory concentrations, MIC) of the seven different Malassezia species to azoles (ketoconazole, itraconazole and voriconazole) and allylamine (terbinafine) was tested. All of the Malassezia yeasts were highly sensitive to the three azole drugs. However, with terbinafine only M. pachydermatis and M. sympodialis were highly sensitive. For the other five Malassezia species the susceptibility to terbinafine was very low for several isolates.

In pityriasis versicolor the lipophilic Malassezia yeasts change from the round yeast cells to the hyphal form. The antifungal activity may be different for the yeast cell and the hyphal form of the organism. In earlier studies we have been able to produce hyphae in Malassezia on human stratum corneum in vitro (12). The yeast was at that time named P. ovale. However, the strain with the highest production of hyphae has later been typed to M. sympodialis. It produced hyphae in 21% of the cells and another M. sympodialis strain produced hyphae in 19% of the cells. The
activity of ketoconazole and itraconazole (13) as well as fluconazole (14) against Malassezia has been studied earlier in this model. In these studies ketoconazole (1 \( \mu \)g/ml) and itraconazole (1 \( \mu \)g/ml) reduced the number of hyphae from 26% to 3% with ketoconazole and to 10% with itraconazole. In the study with fluconazole the production of hyphae was reduced from 25% to 4% when fluconazole (1 \( \mu \)g/ml) was added to the solution with M. sympodialis.

The aim of the present study was to determine: (i) the MIC of R126638 and ketoconazole against the various Malassezia species; and (ii) to investigate the in vitro effect of R126638 and ketoconazole against both the hyphal and yeast cell forms of M. sympodialis.

MATERIALS AND METHODS

Micro-organisms M. pachydermatis CBS 1871, M. furfur CBS 7019, M. sympodialis CBS 7222, M. slooffiae CBS 7956, M. globosa CBS 7966, M. obtusa CBS 7876 and M. restricta CBS 7877 were included in the study. Two other strains known for their ability to produce hyphae on human stratum corneum in vitro: M. sympodialis ATCC 44031 and ATCC 44341 were also included. The strains were maintained on a modified Leeming-Notman agar (7) at 37°C.

R126638 was obtained from Barrier Therapeutics, Geel, Belgium. Ketoconazole was bought from Sigma, St Louis, USA.

Minimal inhibitory concentrations estimated by the agar dilution technique

R126638 and ketoconazole were first dissolved in dimethylformamide (DMF) and then diluted in phosphate buffered saline (PBS, pH 7.4) to give stock solutions containing 100 \( \mu \)g/ml of R126638 and ketoconazole and 20% of DMF. Other stock solutions with lower concentrations were obtained by further dilutions in PBS. From the various stock solutions, dilutions were made directly with the test culture medium. Diagnostic Sensitivity Test agar (DST) (Oxoid, UK) with the addition of Tween 80 (2 ml/l) and glycerol monostearate (2.5 g/l) to obtain concentrations of 0.005–100 \( \mu \)g/ml of R126638 and ketoconazole in the test medium.

Malassezia yeast cells were added on the agar medium, using a Pasteur pipette, in concentrations of 10^7 cells/ml and in a volume of 20 \( \mu \)l. Plates were incubated at 37°C and read after 1, 2 and 3 days. The experiment was performed in duplicate and the MIC was defined as the lowest concentration of the drugs that inhibited growth.

In vitro effect of R126638 and ketoconazole on the filamentous form of M. sympodialis

The method for obtaining stratum corneum has been described in detail earlier (12). Full-thickness skin samples were obtained from plastic surgery on female breasts. Epidermis was peeled off from full-thickness skin by gently heating the skin sample to 65°C, making a break at the basal membrane level. The stratum corneum was obtained by incubating pieces of epidermis in a 0.1% aqueous trypsin solution at 37°C for 2 h, disrupting the desmosomes, then the detached epidermal cells could easily be washed off leaving the stratum corneum intact. Stratum corneum was stored at –70°C until use.

Pieces of stratum corneum (1 cm^2) were placed directly on the culture medium (Leeming-Notman). M. sympodialis (ATCC 44341) in concentrations of 10^5 cells/ml was mixed with R126638 or ketoconazole in concentrations of 0.01, 0.1 and 1 \( \mu \)g/ml and spread over the entire stratum corneum piece. The culture plates were then incubated in a micro-aerophilic environment at 37°C for 6 days. The micro-aerophilic environment was obtained and maintained using the micro-aerophilic system, gas-generating kit from Oxoid (Basingstoke, Hampshire, UK). At the end of the experiment half of the stratum corneum samples were processed for light microscopy and the others were processed for transmission electron microscopy (TEM) and scanning electron microscopy (SEM) (13).

The stratum corneum piece was placed on a glass slide, stained by the periodic acid Schiff reaction (PAS) for light microscopy examination. Twenty-five different fields (\( \times \)1000) were examined and the mean numbers of the percentage of cells producing hyphae were estimated for the various concentrations of R126638, ketoconazole and controls (14).

At the end of the experiment the other half of all stratum corneum samples were immediately fixed at room temperature in 3% glutaraldehyde buffered to pH 7.4 with 0.1 M sodium cacodylate for several days and rinsed in the same buffer supplemented with 7% sucrose. Samples were post-fixed in 1% OsO\(_4\), buffered to pH 7.4 with 0.1 M sodium cacodylate at room temperature for one h. Thereafter, the samples were rinsed in the same buffer and dehydrated in graded series of ethanol. The samples were then embedded in Epon. Ultrathin sections of the cells were stained with uranyl acetate and lead citrate prior to examination in a Philips EM 400 electron microscope (13).

RESULTS

Minimal inhibitory concentration determined by the agar dilution technique

The MICs for R126638 and ketoconazole are shown in Table I. R126638 was more effective than ketoconazole with MICs of 0.01 \( \mu \)g/ml against M. obtusa, M. restricta and all strains of M. sympodialis. However, there was a variation in the sensitivity of the various Malassezia species to both drugs. M. sympodialis, M. restricta, M. obtusa, M. globosa and M. pachydermatis were the species that were most sensitive to R126638 and M. furfur and M. slooffiae were less sensitive. M. globosa and M. slooffiae were the strains with the lower sensitivity to ketoconazole.

<table>
<thead>
<tr>
<th></th>
<th>R126638</th>
<th>Ketoconazole</th>
</tr>
</thead>
<tbody>
<tr>
<td>M. furfur CBS 7019</td>
<td>0.05</td>
<td>0.05</td>
</tr>
<tr>
<td>M. pachydermatis CBS 1871</td>
<td>0.02</td>
<td>0.02</td>
</tr>
<tr>
<td>M. slooffiae CBS 7956</td>
<td>0.05</td>
<td>0.1</td>
</tr>
<tr>
<td>M. globosa CBS 7966</td>
<td>0.02</td>
<td>0.1</td>
</tr>
<tr>
<td>M. obtusa CBS 7876</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>M. restricta CBS 7877</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>M. sympodialis CBS 7222</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>M. sympodialis ATCC 44031</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>M. sympodialis ATCC 44341</td>
<td>0.01</td>
<td>0.01</td>
</tr>
</tbody>
</table>

Acta Derm Venereol 86
In vitro effect of R126638 and ketoconazole on the filamentous form of Malassezia sympodialis

Light microscopy showed that on human stratum corneum in vitro both R126638 and ketoconazole were very effective in reducing the production of hyphae from 15% to 1% with R126638 and to 2% with ketoconazole (Table II). Besides, with higher concentrations of both drugs, many cells were swollen and stained poorly.

Table II. Effect of R126638 and ketoconazole on the production of hyphae in Malassezia sympodialis revealed by light microscopy

<table>
<thead>
<tr>
<th>Substance</th>
<th>Hyphae (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>15</td>
</tr>
<tr>
<td>R126638</td>
<td></td>
</tr>
<tr>
<td>1 µg/ml</td>
<td>1</td>
</tr>
<tr>
<td>0.1 µg/ml</td>
<td>3</td>
</tr>
<tr>
<td>0.01 µg/ml</td>
<td>6</td>
</tr>
<tr>
<td>Ketoconazole</td>
<td></td>
</tr>
<tr>
<td>1 µg/ml</td>
<td>2</td>
</tr>
<tr>
<td>0.1 µg/ml</td>
<td>5</td>
</tr>
<tr>
<td>0.01 µg/ml</td>
<td>9</td>
</tr>
</tbody>
</table>

SEM did not reveal obvious surface differences between untreated cultures and cultures exposed to ketoconazole or R126638 in the concentration range of 0.01 to 1.0 µg/ml. However, TEM showed partial to complete necrosis of the cytoplasmic organelles of M. sympodialis. The fully necrotic cells amounted to 6, 60, 76 and 100% with 0, 0.01, 0.1 and 1 µg/ml, respectively, of R126638 (Fig. 1) and to 6, 41, 62 and 97% with 0, 0.01, 0.1 and 1 µg/ml, respectively, of ketoconazole. There were no differences in morphological changes induced by the azoles between the yeast form and the few remaining budding yeasts.

DISCUSSION

Both R126638 and ketoconazole had a high in vitro activity against the Malassezia yeasts. The two strains of M. sympodialis that easily produce hyphae on human stratum corneum in vitro were very sensitive to R126638. M. sympodialis and M. globosa are the two species that are most commonly isolated from patients with pityriasis versicolor (15), and M. globosa and M. furfur have commonly been cultured from patients with seborrhoeic dermatitis (16).

We described earlier a model that constantly enables the production of hyphae in the Malassezia yeasts (12). This model has been used previously to study the effect of fluconazole (14), ketoconazole and itraconazole (13) on the production of hyphae, as well as on the morphology of the Malassezia yeasts.

In the present study both R126638 and ketoconazole were very effective in blocking the production of hyphae in Malassezia. This was seen both with light- and electron microscopy. The combined SEM and TEM findings confirm earlier observations of the “mummifying” effect of azoles against Malassezia species. A direct necrotic effect has been obtained after azole treatment, showing complete degeneration of internal organelles, but without obvious alterations of the cell periphery. This is clearly shown by light- and electron microscopy. Such a “mummifying” effect of azoles may foster confusion about therapeutic outcome, when therapy is verified by whole mount inspection of the fungus (KOH preparation) as usually done in pityriasis versicolor (17).

R126638 is an interesting new triazole with high activity against the Malassezia yeasts, the yeasts involved in pityriasis versicolor and seborrhoeic dermatitis. In conclusion, this model for in vitro production of hyphae in Malassezia is valuable for screening the activity of antifungal agents against the filamentous form of these yeasts.

ACKNOWLEDGEMENT

This work was partly supported (J.A.) by a grant from IWT-Vlaanderen (grant no. 030023).
REFERENCES