

Evaluation of Single Nucleotide Variants in Ethiopian Patients with Atopic Dermatitis

Samina ASAD¹, Isabel TAPIA-PÁEZ^{1,2}, Andrea Montano MONTES¹, Carl-Fredrik WAHLGREN¹, Kassahun D. BILCHA³, Magnus NORDENSKJÖLD² and Maria BRADLEY^{1,2}

¹Dermatology Unit, Department of Medicine Solna, Karolinska Institutet, ²Center for Molecular Medicine, Department of Molecular Medicine and Surgery, Karolinska Institutet, SE-171 76, Stockholm, Sweden, and ³Department of Dermatovenereology, Faculty of Medicine, Gondar University, Gondar, Ethiopia. E-mail: samina.asad@ki.se

Accepted Sep 23, 2018; Epub ahead of print Sep 24, 2018

Atopic dermatitis (AD) is an inflammatory skin disorder characterized by pruritus and dryness of the skin in typical locations. The incidence rate of AD varies worldwide. In Europe the prevalence is up to 20% in children, while reports in Ethiopia vary between 1% and 2% in rural areas and 19% in urban areas (1, 2).

The major risk gene for AD in Europeans is the filaggrin (*FLG*) gene (1q21), where a number of null-mutations as well as variations in gene dose are associated with disease susceptibility (2). *FLG* mutations have also been identified in the Asian population, although these mutations seem to be more population- or family-specific (2). Little is known about AD susceptibility genes in the African population. Our previous studies indicate a low prevalence of *FLG* null-mutations in Ethiopian cases of AD (1). This is in line with studies performed in the South African AmaXhosa population, suggesting that, in the African population, genes other than *FLG* may be involved in the development of AD (3).

In searching for risk genes for AD in the Ethiopian population, we previously performed whole-exome sequencing (WES) of 22 Ethiopian patients with AD/ichthyosis vulgaris in whom several genetic polymorphisms were detected (1).

This study aimed to elucidate whether some of the single-nucleotide variants (SNVs) identified by Taylan et al. are associated with AD susceptibility in our Ethiopian AD cohort, consisting of 184 cases of AD and 186 healthy controls (Table I) including the 22 previously sequenced individuals (1).

METHODS

All study participants gave written or oral informed consent, which complies with the principles of the Declaration of Helsinki. This study was approved by the Ethics Review Board of the University of Gondar, and the regional ethics committee, Karolinska Institutet, Sweden.

All patients were diagnosed with AD according to the UK Working Party's diagnostic criteria (2) and answered questionnaires

regarding other atopic manifestations (1). The samples were obtained from ALERT Dermatology Hospital, Tikur Anbessa Hospital and Gondar University Hospital. We chose to genotype gene variants in: (i) epidermal differentiation complex (EDC) genes; cornulin (*CRNN*) (4) and hornerin (*HRNR*) (5); (ii) non-EDC genes that had either previously been associated with AD or candidate genes with a known function in the skin; (iii) gene variants that more than one individual was carrying in the WES. Based on this strategy we genotyped 38 SNVs using TaqMan (Applied Biosystems Inc., Foster City, CA, USA) in the following 10 genes; transglutaminase (*TGM*) 1, 3 and 5 (6), desmoglein (*DSG*) 1 and 3–4 (7), a variant in *C11orf30* encoding for the EMSY protein (8), *SPINK5* (11), hornerin (*HRNR*) (5) and cornulin (*CRNN*) (4). Primers/probes were obtained from Thermo-Fisher Scientific Inc., (Foster City, CA, USA). Genotyping indicated that 28 SNVs were non-polymorphic or did not detect minor allele homozygotes in our material (Table SI¹). For the remaining 10 SNVs (Table II and Table SII¹), association with AD was calculated using χ^2 tests on both genotype and allele frequencies. Fisher's exact test was performed in instances where the counts were lower than 5. R program v.2.12.2 was used to calculate logistic-regression using generalized linear modelling between patients and controls. SNV frequencies were checked to follow Hardy-Weinberg equilibrium.

RESULTS

The genotyping success and concordance rates for SNVs showing 3 genotypes was above 90%. Furthermore, all successfully genotyped SNVs (showing all 3 genotypes) correlated with the previous whole exome-sequence results.

In our WES study, 12 SNVs were identified in the *HRNR* gene, which is a component of the epidermal cornified cell envelope thought to share similar features with filaggrin and demonstrated to have reduced expression in the epidermis of patients with AD (5). In the *HRNR* gene, only 3 SNVs demonstrated 3 genotypes and were analysed further. Two SNVs were analysed in *CRNN*, which is a marker of late epidermal differentiation and down-regulated in AD skin (4). However, these SNVs were non-polymorphic in our study material and discarded from further studies. In addition, SNVs were genotyped in *DSG1-4* and *TGM1*, 3 and 5, all of which have important roles in forming the epidermis and are associated with altered expression in AD skin (6, 7). One SNV located in the *c30orf11* region, which has been significantly associated with AD in previous studies, was also analysed (8). In the *SPINK5* gene, 7 SNVs were genotyped. *SPINK5* (5q32) encodes for the

Table I. Ethiopian cases and controls

	Cases (n = 184)	Controls (n = 186)
Female, %	46	51
Age, median; range	17; 2 months–45 years	12; 5–60 years
Early age of onset (<2 years old), %	68	n.a.
Moderate AD (SCORAD 15–40), %	30	n.a.
Severe AD (SCORAD >40), %	70	n.a.

AD: atopic dermatitis; SCORAD: SCORing Atopic Dermatitis; n.a.: not applicable.

¹<https://www.medicaljournals.se/acta/content/abstract/10.2340/00015555-3051>

Table II. Minor allele frequencies (MAF) in our cases-control material and for the African population in Exac-and GnomAD databases

SNV name	Gene name	Minor allele	MAF AD patients (n = 184)	MAF controls (n = 186)	Exac African	GnomAD African	p-value
HRNR2 (HRNR:NM_001009931:exon3:c.C2686T:p.R896C)	HRNR	G	0.05	0.07	–	–	0.52
Rs111789738	HRNR	T	0.096	0.086	0.024	0.022	0.88
Rs77898999	HRNR	G	0.023	0.02	0.016	0.015	0.34
SPINK5_6 (SPINK5:NM_001127698:exon7:c.G532A:p.E178K,SPINK5:NM_001127699:exon7:c.G532A:p.E178K,SPINK5:NM_006846:exon7:c.G532A:p.E178K)	SPINK5	G	0.062	0.07	–	–	0.92
Rs2303063	SPINK5	G	0.34	0.4	0.24	0.24	0.07
Rs2303067	SPINK5	G	0.35	0.34	0.24	0.24	0.24
Rs3188691	SPINK5	A	0.063	0.068	0.025	0.023	0.27
Rs114998364	TGM3	G	0.062	0.051	0.031	0.028	0.57
Rs36040686	DSG4	C	0.077	0.065	0.046	0.033	0.63
Rs140750904	DSG4	G	0.039	0.023	0	0.00012	0.54

p-values for the whole Ethiopian cases-control cohort for all successfully genotyped single-nucleotide variants (SNVs). AD: atopic dermatitis.

skin barrier protein *Lympho-epithelial Kazal-type-related inhibitor* (LEKTI), which plays a major role in controlling the proteolytic processing of profilaggrin into FLG monomers. Loss-of-function mutations of *SPINK5* cause the rare autosomal recessive disorder Netherton syndrome characterized by congenital ichthyosis as well as manifestations overlapping with AD. SNVs in this gene have previously shown modest association with AD in European and Asian populations (9). When association was calculated only between patients with severe AD and controls, significant association was observed for both rs230363 ($p=0.023$) and rs2303067 ($p=0.025$) in *SPINK5*. These findings are in line with previous studies in European and Asian populations (9). This prompted us to perform immunohistochemistry with the *SPINK5* antibody (Novus Biologicals, Abingdon Oxon, UK), on skin biopsies of Ethiopian patients with AD. Similar staining signals were indicated in a rs230363 homozygous wild-type carrier a homozygous minor allele-carrier and a heterozygous carrier (Fig. S1¹). However, we did not find any association when logistic regression was calculated. Association was also tested in all SNVs for age at onset (ages <2, 5 and 8 years, respectively) vs. controls and for AD severity measured by SCORing Atopic Dermatitis (SCORAD).

DISCUSSION

In our study, most SNVs had low minor allele frequencies (MAF) corresponding to frequencies in the African population in the online databases ExAc and GnomAD (Table II and Table S1¹) and, therefore, since our cohort is small, we may be limited to identify all 3 genotypes for many of the SNVs. Although it may be concluded that, due to low MAF, none of the variants would have a large effect on risk of AD. On the other hand, since most of the genes analysed are known to have altered expression in patients with AD compared with healthy controls, a dose-dependent or change of expression of the genes in the Ethiopian population that lead to AD cannot be ruled out.

Bearing in mind that *FLG* is not associated with AD in the Ethiopian population and no other major disease-susceptibility genes have been identified so far these re-

sults indicate that more genetic studies need to be carried out in order to understand the genetic background of AD in the Ethiopian population.

ACKNOWLEDGEMENTS

All patients and healthy control individuals attending the Department of Dermatovenereology, Faculty of Medicine, Gondar University, Gondar, Ethiopia. Supported by Edward Welander Foundation (2206) and the Swedish Asthma and Allergy Association's Research Foundation (2013040) and through the regional agreement on medical training and clinical research (ALF) between Stockholm County Council and Karolinska Institutet.

The authors have no conflicts of interest to declare.

REFERENCES

- Taylan F, Nilsson D, Asad S, Lieden A, Wahlgren CF, Winge MC, et al. Whole-exome sequencing of Ethiopian patients with ichthyosis vulgaris and atopic dermatitis. *J Allergy Clin Immunol* 2015; 136: 507–509.
- Brown SJ, Kroboth K, Sandilands A, Campbell LE, Pohler E, Kezic S, et al. Intragenic copy number variation within filaggrin contributes to the risk of atopic dermatitis with a dose-dependent effect. *J Invest Dermatol* 2012; 132: 98–104.
- Thawer-Esmail F, Jakasa I, Todd G, Wen Y, Brown SJ, Kroboth K, et al. South African amaXhosa patients with atopic dermatitis have decreased levels of filaggrin breakdown products but no loss-of-function mutations in filaggrin. *J Allergy Clin Immunol* 2014; 133: 280–282.
- Lieden A, Ekelund E, Kuo IC, Kockum I, Huang CH, Mallbris L, et al. Cornulin, a marker of late epidermal differentiation, is down-regulated in eczema. *Allergy* 2009; 64: 304–311.
- Henry J, Hsu CY, Haftek M, Nachat R, de Koning HD, Gardinal-Galera I, et al. Hornerin is a component of the epidermal cornified cell envelopes. *FASEB J* 2011; 25: 1567–1576.
- Lieden A, Winge MC, Saaf A, Kockum I, Ekelund E, Rodriguez E, et al. Genetic variation in the epidermal transglutaminase genes is not associated with atopic dermatitis. *PLoS One* 2012; 7: e49694.
- Samuelov L, Sarig O, Harmon RM, Rapaport D, Ishida-Yamamoto A, Isakov O, et al. Desmoglein 1 deficiency results in severe dermatitis, multiple allergies and metabolic wasting. *Nature Genetics* 2013; 45: 1244–1248.
- O'Regan GM, Campbell LE, Cordell HJ, Irvine AD, McLean WH, Brown SJ. Chromosome 11q13.5 variant associated with childhood eczema: an effect supplementary to filaggrin mutations. *J Allergy Clin Immunol* 2010; 125: 170–174.
- Dezman K, Korosec P, Rupnik H, Rijavec M. *SPINK5* is associated with early-onset and *CHI3L1* with late-onset atopic dermatitis. *Int J Immunogenetics* 2017; 44: 212–218.