Peripheral NMDA Receptor/NO System Blockage Inhibits Itch Responses Induced by Chloroquine in Mice

Nazgol-Sadat HADDADI^{1,2,4}, Arash FOROUTAN^{1,3}, Sattar OSTADHADI^{1,2,5}, Ehsan AZIMI⁶, Nastaran RAHIMI^{1,2}, Mehdi NATEGHPOUR⁴, Ethan A. LERNER⁶ and Ahmad Reza DEHPOUR¹⁻³

¹Experimental Medicine Research Center, ²Department of Pharmacology, School of Medicine, ³Faculty of Pharmacy, ⁴Department of Medical Parasitology and Mycology, School of Public Health, ⁵Brain and Spinal Cord Injury Research Center, Neuroscience Institute, Tehran University of Medical Sciences, Tehran, Iran and ⁶Department of Dermatology, Cutaneous Biology Research Center, Massachusetts General Hospital, Boston, USA

Intradermal administration of chloroquine (CQ) provokes scratching behavior in mice. Chloroguine-induced itch is histamine-independent and we have reported that the nitric oxide (NO)/cyclic guanosine monophosphate (cGMP) pathway is involved in CQ-induced scratching behavior in mice. Previous studies have demonstrated that activation of N-methyl-d-aspartate receptors (NMDARs) induces NO production. Here we show that NMDAR antagonists significantly decrease CQ-induced scratching in mice while a non-effective dose of an NMDAR agonist potentiates the scratching behavior provoked by sub-effective doses of CQ. In contrast, combined pre-treatment with sub-effective doses of an NMDAR antagonist, MK-801, and the NO synthase inhibitor, L-N-nitro arginine methyl ester (L-NAME), decreases CQ-induced scratching behavior. While intradermal administration of CQ significantly increases the concentration of intradermal nitrite, the end product of NO metabolism, effective doses of intraperitoneal and intradermal MK-801 significantly decrease intradermal nitrite levels. Likewise, administration of an effective dose of L-NAME significantly decreases CQ-induced nitrite production. We conclude that the NMDA/NO pathway in the skin modulates CQinduced scratching behavior.

Key words: itch; chloroquine; *N*-methyl-D-aspartate receptor (NMDA) antagonists; nitric oxide; mice.

Accepted Jan 24, 2017; Epub ahead of print Jan 25, 2017

Acta Dem Venereol 2017; 97: 571-577.

Corr: Ahmad Reza Dehpour, Department of Pharmacology, School of Medicine, Tehran University of Medical Sciences, Poorsina St., Enghelab Ave., PO Box: 13145-784, Tehran, Iran. E-mail: dehpour@yahoo.com

Chloroquine (CQ) has been used to treat malaria, certain viral infections (1) and a number of autoimmune diseases including rheumatoid arthritis (2) and systemic lupus erythematosus (3). Chloroquine also induces itch in humans and scratching in mice (4, 5) via a histamine-independent pathway linked to activation of Mrgprs (6). Chloroquine activates human MRGPRX1 and the homologous mouse receptor, MrgprA3 (6). As expected, CQ-induced scratching behavior is diminished in Mrgpr cluster knockout mice, which lack MrgprA3 (7). In addition, the kappa-opioid agonist, nalfurafine, has been shown to decrease CO-induced scratching behavior in mice (8). MrgprA3 coupling to $G_{R_{H}}$ modulates TRPA1 to result in itch (6, 9). $G_{g_{y}}$ modulates several ion channels via direct binding (10). The N-methyl-daspartate receptor (NMDAR) is a non-selective cation ion channel (11). NMDAR is implicated in synaptic plasticity, memory (11, 12) and pain (13, 14). The administration of NMDA or glutamate to the rat hind paw evokes nociceptive behaviors that are decreased by the injection of NMDAR antagonists in the periphery (14, 15). Peripherally-acting NMDAR antagonists have been used for inflammatory and visceral pain and seem to be devoid of CNS side effects (16) while oral administration of gabapentin and pregabalin act centrally and are used to treat the neuropathic itch associated with brachioradial and post-herpetic pruritus (17–21).

Intrathecal injection of NMDA induces scratching behavior in mice (22) while NMDAR antagonists suppress such behavior (17, 23). The physiology is analogous in rats, where intrathecal administration of the NMDAR antagonists ketamine and kynurenic acid decrease scratching behavior induced by intradermal injection of the serotonin derivative 5-methoxytryptamine (23). Intrathecal application of the NMDAR antagonists dizocilpine (MK-801) or D (-) -2-amino- 5-phosphonovalerate (APV) prevented intracutaneous histamineinduced expansion of mechanical receptive fields in rats (24). Separately, it has been shown that administration of a nitric oxide (NO) donor to skin enhances substance P-induced itch in mice (25), while serotonin-induced itch was inhibited by systemic nitric oxide synthase (NOS) inhibitors (26).

Taken together, previous reports have revealed that activation of the NMDA/NO pathway is associated with pain (27) while inhibition of this pathway decreases pain and itch (23, 25, 27). Activation of this pathway is also associated with increased NO production (28), which itself is linked to cGMP (29). NO production is associated with itch while inhibition of NOS suppresses itch. We reported recently that the NO/cGMP pathway participates in CQ-induced itch (30). Here we extend these findings with the demonstration that the NMDA/ NO pathway participates in CQ-induced scratching behavior.

METHODS

Animals and material

Male NMRI mice, an outbred strain, (Pasteur Institute, Tehran, Iran) of 5-6 weeks of age and weight range of 23-30 g were used.

Animals were maintained in appropriate facilities with respect to temperature (23-25°C) and light (lights on from 08:00 AM to 08:00 PM) and had free access to food or water (31).

All of the operational guidelines in the housing, routine husbandry, handling, and experimental procedures were approved by the committee for animal ethics and experiments at Tehran University of Medical Sciences, Tehran, Iran.

Chloroquine bisphosphate (Pubchem CID 64927) was a gift from Pars Darou Pharmaceutical Company (Tehran, Iran).

Loratadine (Pubchem CID 3957), magnesium sulfate (Pubchem CID 24083), ketamine hydrochloride solution (Pubchem CID 15851 - CAS Number: 1867-66-9), MK-801 (Pubchem CID 1207), N-methyl-D-aspartic acid (NMDA) (Pubchem CID 22880), N-nitro-L-arginine methyl ester (L-NAME) (Pubchem CID 39836) and vanadium chloride (Pubchem CID 62647) were purchased from Sigma, St. Louis, MO, USA. Lysis buffer was purchased from Abcam, Cambridge, MA, USA.

All drugs except loratadine were prepared freshly for use by dissolving in physiological saline. Loratadine was dissolved in phosphate-buffered saline (PBS).

Drug administration

Loratadine, a second-generation histamine type 1 (H1) receptor antagonist, was injected intraperitoneally (IP) at a dose of 10 mg/ kg 30 min before intradermal (ID) injection of 400 µg CQ.

Magnesium sulfate (MgSO₄) was administered IP at doses of 5 and 20 mg/kg 30 min before ID injection of 400 µg CQ in volume of 50 µl per site. Magnesium ions block the NMDA type of ionotropic glutamate receptors (ion GluRs) (32).

We targeted the glutamate site of NMDA receptors by the glutamate channel blockers ketamine and MK-801, at doses of 2, 5 mg/kg and 0.1, 0.25 mg/kg, respectively injected IP 45 min beforehand (32, 33). NMDA was injected intraperitoneally (IP) at a dose of 75 mg/kg, 30 min before ID administration of CQ at a subeffective dose of 200 µg/site (32, 33). MK-801 was administered ID at a dose of 10 nmol simultaneously with CQ 400 µg/site (34).

We recently reported that IP injection of 3 and 10 mg/kg of L-NAME significantly (p < 0.001) reduces CQ 400 µg-induced scratching behavior. We also demonstrated that L-NAME at a dose of 1 mg/kg is considered sub-effective and does not significantly inhibit CO 400 µg-induced scratching behavior (30). For the current study, sub-effective doses of intraperitoneal MK-801 (0.1 mg/kg) and L-NMAE (1 mg/kg) were injected 45 and 30 min before CQ 400 µg, respectively.

Behavioral experiments

All animals were habituated in an acrylic box $(10 \times 10 \times 13 \text{ cm})$ at 23° C±1 for 1 h before behavioral experiments. A small amount of bedding was placed in the box. Hair was removed from the rostral back of the mice by depilatory cream. After two days, ID injections were delivered to the shaved area in a volume of 50 µl per site and each mouse was used once. The mice were removed briefly from the box for injections, returned to the same box after injections; and the behavior was recorded using a video camera in unmanned conditions to avoid distraction. The video was played back to quantify the scratching bouts directed at the site of injection. Each scratching bout is initiated by lifting of the hind paw to the area of injection, and ended by returning of the hind paw to the floor or to the mouth.

Open-field locomotor activity

NMDAR antagonists are associated with dissociative symptoms. To rule out the possibility of such effects on the ambulatory behavior of mice, an open-field test (35) was performed to evaluate the effect of NMDAR antagonists on the motor activity of mice. The apparatus consisted of a wooden box measuring $40 \times 60 \times 50$ cm. The floor of the arena was divided into 12 equal squares. The animals were gently placed in the center of the field, and the number of squares crossed with all paws (crossing) was counted in a 6-min session. Although loratadine is a non-sedative H1 receptor antagonist, we performed an open-field test with loratadine to rule out its possible effects on locomotor activity of the mice.

Measurement of nitric oxide levels in skin tissue

Mice were sacrificed by cervical dislocation and the rostral skin (the site of injection) was removed 5, 15, 25 and 35 min after ID injection of CO and saline. Next, we evaluated the changes in nitrite concentration after injection of an effective dose of L-NAME (10 mg/kg, IP) and 15 min after injection of CQ (400 µg, ID) (the time of maximum scratching behavior and nitrite concentration). We also evaluated the effects of MK-801 (0.25 mg/kg, IP and 10 nmol/site, ID) on the nitrite level 15 min after CQ injection (400 µg, ID).

NO metabolite, nitrite, was measured in the homogenized supernatant samples using the Griess reaction (36). One tenth ml of washout samples were pipetted into a 96-well micro titer plate, then 0.1 ml of Griess reagents containing 2.5% w/v sulphanilamide and 2.5% N-(1-naphthyl) ethylenediamine hydrochloride were added and incubated at room temperature for 15 min to allow color development. One tenth ml of 5% w/v vanadium chloride was added and incubated at 37°C for 45 min. Nitrite concentration was calculated using ELISA and the results were expressed as pmol/mg.

Data analysis

Data were processed (GraphPad Prism 6.0 graphing and statistics software) by one-way or two-way analysis of variance (ANOVA) along with Dunnett's test or Tukey's multiple comparisons tests. The *t*-test was performed for some data analyses. In all the experiments, p < 0.05 was considered significant. Data are presented as mean ± standard error of the mean (SEM).



Fig. 1. The effect of loratadine on chloroquine (CQ)-induced scratching behavior. Administration of a non-sedative H1 antagonist, loratadine, (10 mg/kg, intraperitoneally (IP)) 30 min before 400 µg CQ (intradermally (ID)) does not significantly reduce CQ-induced scratching behavior (p = 0.3625). Values are expressed as mean ± SEM (n = 8) and were analyzed using a *t*-test. p > 0.05.

RESULTS

Chloroquine induces histamine-independent scratching behavior in NMRI mice

Previous studies with C57BL/6 mice have demonstrated that CQ-induced itch is histamine-independent (6). To confirm these findings, we evaluated the effect of loratadine, a non-sedating H1 receptor antagonist, on CQ-induced scratching behavior in NMRI mice. The administration of loratadine (10 mg/kg, IP) before CQ (400 µg/site) did not significantly decrease the pruritic behavior (p > 0.05) (**Fig. 1**). Consistent with its non-sedating activity, loratadine (10 mg/kg, IP) had no significant effect on the locomotor activity of mice in the open-field test (p > 0.05; data not shown) (37).

NMDAR antagonists decrease chloroquine-induced scratching behavior

Previous studies have revealed that NMDA induces itch (22) while NMDAR antagonists suppress itch (17, 23). We thus asked if NMDAR antagonists would decrease scratching behavior from CQ. Mice were pretreated with MgSO₄ (5 and 20 mg/kg), ketamine (2 and 5 mg/kg), and MK-801 (0.1 and 0.25 mg/kg) before intradermal CQ injection (400 μ g/site). Chloroquine-induced scratching behavior was significantly decreased when effective doses of MgSO₄ (20 mg/kg), ketamine (5 mg/kg) or MK-801 (0.25 mg/kg) were administered intraperitoneally (F (7, 56) = 11.75, *p* < 0.0001) (**Fig. 2**). MK-801, ketamine and magnesium sulphate had no significant effect on the locomotor activity of mice in the open field test (F (3,



CQ 400 µg (ID)

Fig. 2. Systemic administration of NMDA receptor antagonists decreases chloroquine (CQ)-induced scratching behavior. $MgSO_4$ (5 and 20 mg/kg, intraperitoneally (IP)) was administered 30 min before 400 µg CQ (intradermally (ID)) in volume of 50 µl per site. MK-801 (0.1 and 0.25 mg/kg, IP) and ketamine (2 and 5 mg/kg, IP) were administered 45 min before CQ 400 µg (ID). Values are expressed as mean ± SEM (n = 8) and were analyzed using a one-way ANOVA followed by Dunnett's test. ****p < 0.0001.



Fig. 3. Intradermal MK-801 (10 nmol/site) significantly decreases chloroquine (CQ)-induced scratching behavior. MK-801 (10 nmol/site, intradermally (ID)) *per se* did not have any significant pruritic effects. Values are expressed as mean \pm SEM (n = 8) and were analyzed using a two-way ANOVA type III followed by Tukey's multiple comparisons test. ****p < 0.0001.

28) = 0.83, p > 0.05; data not shown) at these concentrations. **Fig. 3** shows the effect of MK-801 (10 nmol/site) on scratching induced by CQ 400 µg/site. A two-way ANOVA type III showed the effect of intradermal MK-801 (F (1, 28) = 0.0218, p > 0.05), the effect of CQ 400 µg/site (F (1, 28) = 53.9007, p < 0.0001) and the effect of intradermal MK801 × CQ 400 µg/site interaction (F (1, 28) = 27.4496, p < 0.0001). Intradermal MK-801 (10 nmol/site) decreased CQ-induced scratching levels to those of mice treated with saline alone.

NMDA potentiates CQ-induced scratching behavior

As NMDA can itch (22), we asked if NMDAR agonists could allow for itch to be induced by a sub-itch-inducing



CQ 200 μg (ID)

Fig. 4. NMDA potentiates chloroquine (CQ)-induced scratching behavior. Pretreatment with the NMDA agonist (NMDA, 75 mg/kg, intraperitoneally (IP)) 30 min before administration of CQ 200 µg (intradermally (ID)) potentiated the scratching responses (p = 0.0103). NMDA (75 mg/kg, IP) per se does not have any significant pruritic effects. Values are expressed as mean ± SEM (n = 8) and were analyzed using two-way ANOVA type III followed by Tukey's multiple comparisons test.*p < 0.05.

dose of CQ. **Fig. 4** demonstrates the effect of a NMDAR agonist on CQ-induced scratching behavior. Mice were pretreated with NMDA (75 mg/kg, IP) in advance of sub-pruritic intradermal CQ (200 μ g/site) injection. A two-way ANOVA type III showed that CQ (200 μ g/site) × NMDA (75 mg/kg, IP) injection induces significant scratching behavior (F (1, 28) = 5.0501, *p*<0.05). No significant pruritic effect was observed with NMDA (75 mg/kg, IP) injection (F (1, 28) = 0.0501, *p*>0.05). We conclude that NMDA per se does not induce considerable scratching behavior, is significantly potentiated in the presence of NMDA (75 mg/kg, IP).

Combined sub-effective doses of MK-801 and L-NAME decrease chloroquine-induced scratching behavior

As NMDAR antagonists and NOS antagonists can suppress itch (17, 23, 25), we next asked if low, thus individually sub-effective doses of such compounds, would have an additive effect and decrease CQ-induced scratching. Sub-effective doses of L-NAME and MK-801 alone had no significant effect on CQ-induced scratching behavior, the combination of sub-effective doses of L-NAME (1 mg/kg) and MK-801 (0.1 mg/kg), significantly decreased CQ-induced scratching behavior (F (3, 28) = 19.50, p < 0.0001) (Fig. 5).

Intradermal administration of chloroquine increases the concentration of nitrite

Given that NO is associated with itch (38), we next asked if administration of CQ would increase the level of nitrite, the end product of NO metabolism in the skin. Intradermal CQ (400 μ g/site) induced scratching behavior with



CQ 400 µg (ID)

Fig. 5. The effect of MK-801 and L-NAME on chloroquine (CQ)-induced scratching behavior. Combined treatment with sub-effective doses of MK-801 (0.1 mg/kg, intraperitoneally (IP)) 45 min before and L-NAME (1 mg/kg, IP) 30 min before administration of CQ 400 µg intradermally (ID) significantly reduced CQ-induced scratching responses (p= 0.0002). Values are expressed as the mean±SEM (n=8). ***p < 0.001.



Fig. 6. Chloroquine (CQ)-induced scratching behavior and production of nitrite in mouse skin. (A) The time-course of scratching activity following intradermal injection of CQ 400 µg/site (n = 8). (B) The time-course of nitrite production following intradermal injection of CQ 400 µg/ site. Skin injected with chloroquine was isolated 5, 15, 25 and 35 min after CQ injection. The concentration of nitrite is increased compared to the saline group in 5 (p = 0.0004) and 15 min (p < 0.0001) in the site of injection. Each group consisted of 8 animals. Values are expressed as mean ± SEM (n = 8) and were analyzed using two-way ANOVA followed by Tukey's multiple comparisons test. +++p < 0.001, ****p < 0.0001.

maximum scratching behavior at 10 to 20 min after injection (**Fig. 6**A). A two-way ANOVA showed the effect of CQ 400 µg/site (F (1, 56) = 32.28, p < 0.0001), the effect of time (F (3, 56) = 37.57, p < 0.0001) and the effect of CQ 400 µg/site × time interaction (F (3, 56) = 8.212, p = 0.0001). Intradermal CQ significantly increased the concentration of nitrite compared to the saline group in 5 and 15 min in the site of injection with maximum concentration at 15 min after CQ administration (Fig. 6B). L-NAME (10 mg/kg, IP) and MK-801 (0.25 mg/kg, IP and 10 nm/site, ID) significantly decreased the nitrite levels 15 min after CQ 400 µg/site injection (F (4, 35) = 27.72, p < 0.0001) (**Fig. 7**).

DISCUSSION

The NMDA/NO pathway has been associated with pain (27) while inhibition of this pathway decreases pain and itch (23, 25–27). Activation of this pathway is also associated with increased production of NO (28). NO is associated with itch while inhibition of NOS suppresses itch (25, 26, 30). We reported recently that the NO/cGMP

ActaDV

Advances in dermatology and venereology





CQ 400 µg (ID)

Fig. 7. L-NAME and MK-801 significantly decrease chloroquine-(CQ) induced nitrite levels in the skin. Mice were treated with L-NAME and MK-801 and nitrite levels were measured 15 min after CQ (400 μ g/site) injection. Values are expressed as mean \pm SEM (n=8). ####p < 0.0001and ****p < 0.0001 compared with saline and CQ alone, respectively.

pathway participates in CQ-induced itch (30). The data here extend these findings. These data demonstrate that the NMDA/NO pathway participates in CQ-induced scratching behavior. These data also reveal that CQinduced scratching behavior is significantly decreased by the NMDA receptor antagonists MK-801, ketamine and MgSO₄. Conversely, CQ-induced scratching is potentiated in the presence of NMDA. Consistent with these findings, CQ was found to promote the generation of nitrite, a proxy for NO production, the production of which was decreased after intradermal and intraperitoneal injection of the NMDAR antagonist MK-801. The data thus support a role for peripheral NMDA receptors together with the NO pathway in CQ-induced itch.

CQ activates MrgprA3 and coupling of this receptor to TRPA1 via $G_{\beta\gamma}$ is necessary for CQ-induced itch (39). Chloroquine does not induce itch in TRPA1 knockout mice (9). Previous investigations have described interactions between glutamatergic signaling and TRPA1 channels (27). Intraplantar injection of formalin produces TRPA1-dependent biphasic nociceptive responses in rodents. Formalin directly activates TRPA1 channels on primary afferents followed by the release of inflammatory mediators (40). Glutamate, an endogenous NMDAR ligand (41), is also increased in the glabrous skin of the rat hind paw after formalin administration (42). TRPA1 channels may mediate CQ-induced itch by activating the glutamatergic system and NMDA receptors. Previous studies have demonstrated that more than 80% of the mouse dorsal root ganglion neurons responding to chloroquine, are immunopositive for vesicular glutamate transporter type 2 (VGLUT2) proposing a role for glutamate in CO-induced itch (43). Glutamate binds the NMDAR, leading to ion channel opening and release of Mg^{2+} (44) which allows for an influx of Ca^{2+} , stimulation of nNOS activity and subsequent NO production

(28). In addition to pain (27, 29, 45), the NMDA/NO synthase pathway has been implicated in modulation of many other behavioral processes including depression, learning, anxiety, aggression and locomotion (46–50). Our findings in the periphery, that activation of NMDA receptors following CQ injection stimulates NO production in skin, complement what has been found centrally.

NO functions as a neurotransmitter in itch (25, 26, 38, 51). Similar to our findings with CO, previous studies have shown that NO enhances substance P-induced itch in mice (25). In addition, NO production is increased in the skin of patients with pruritic skin diseases such as atopic dermatitis and psoriasis (52, 53). Although, the exact cite of action of CO-induced NO is not clear, it has been suggested that NO may directly activate primary sensory neurons (54). In addition, NO can also bind to TRPA1 and modulate its function by cysteine S-nitrosylation (55, 56). Keratinocytes and primary sensory neurons express NMDAR, nNOS and TRPA1 (34, 57-63) while CQ activates MrgprA3 (6). Sensory neurons may thus not be the only source of NO following intradermal CO injection. CO also induces mast cell degranulation, (64, 65) releasing mediators that interact with afferent nerve fibers in the skin.

In conclusion, our findings reveal that peripheral NMDA receptors and the production of NO in skin may be part of the pathway that underlies CQ-induced itch. Targeting peripheral NMDA receptors may be of benefit in the treatment of histamine-independent itch.

ACKNOWLEDGMENTS

This study was supported by Experimental Medicine Research Center, Tehran University of Medical Sciences, Tehran, Iran (Grant number: 94-02-30-29012). Research reported in this publication was supported by the National Institute of Arthritis and Musculoskeletal and Skin Diseases of the National Institutes of Health under Award Numbers R01AR057744 and R21AR067399 to EAL. EA is the recipient of a grant from the National Eczema Association. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

The authors declare no conflicts of interest.

REFERENCES

- Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases. Lancet Infect Dis 2003; 3: 722–727.
- Romanelli F, Smith KM, Hoven AD. Chloroquine and hydroxychloroquine as inhibitors of human immunodeficiency virus (HIV-1) activity. Curr Pharm Des 2004; 10: 2643–2648.
- Borba E, Turrini-Filho J, Kuruma KA, Bertola C, Pedalini ME, Lorenzi M, et al. Chloroquine gestational use in systemic lupus erythematosus: assessing the risk of child ototoxicity by pure tone audiometry. Lupus 2004; 13: 223–227.
- Green AD, Young KK, Lehto SG, Smith SB, Mogil JS. Influence of genotype, dose and sex on pruritogen-induced scratching behavior in the mouse. Pain 2006; 124: 50–58.
- 5. Sowunmi A, Walker O, Salako L. Pruritus and antimalarial

drugs in Africans. Lancet 1989; 334: 213.

- 6. Liu Q, Tang Z, Surdenikova L, Kim S, Patel KN, Kim A, et al. Sensory neuron-specific GPCR Mrgprs are itch receptors mediating chloroquine-induced pruritus. Cell 2009; 139: 1353-1365.
- 7. Bautista DM, Wilson SR, Hoon MA. Why we scratch an itch: the molecules, cells and circuits of itch. Nat Neurosci 2014; 17: 175-182.
- 8. Akiyama T, Carstens MI, Piecha D, Steppan S, Carstens E. Nalfurafine suppresses pruritogen-and touch-evoked scratching behavior in models of acute and chronic itch in mice. Acta Derm Venereol 2015; 95: 147-150.
- 9. Wilson SR, Gerhold KA, Bifolck-Fisher A, Liu Q, Patel KN, Dong X, et al. TRPA1 is required for histamine-independent, Mas-related G protein-coupled receptor-mediated itch. Nat Neurosci 2011; 14: 595-602.
- 10. Dascal N. Ion-channel regulation by G proteins. Trends Endocrinol Metab 2001; 12: 391-398.
- 11. McBain C, Mayer M. N-methyl-D-aspartic acid receptor structure and function. Physiol Rev 1994; 74: 723-760.
- 12. Castellano C, Cestari V, Ciamei A. NMDA receptors and learning and memory processes. Curr Drug Targets 2001; 2: 273-283.
- 13. Horvath G, Joo G, Dobos I, Klimscha W, Toth G, Benedek G. The synergistic antinociceptive interactions of endomorphin-1 with dexmedetomidine and/or S (+)-ketamine in rats. Anesth Analg 2001; 93; 1018-1024.
- 14. Petrenko AB, Yamakura T, Baba H, Shimoji K. The role of N-methyl-D-aspartate (NMDA) receptors in pain: a review. Anesth Analg 2003; 97: 1108-1116.
- 15. Zhou S, Bonasera L, Carlton SM. Peripheral administration of NMDA, AMPA or KA results in pain behaviors in rats. Neuroreport 1996; 7: 895-900.
- 16. Parsons CG. NMDA receptors as targets for drug action in neuropathic pain. Eur J Pharmacol 2001; 429: 71-78.
- 17. Cevikbas F, Steinhoff M, Ikoma A. Role of spinal neurotransmitter receptors in itch: new insights into therapies and drug development. CNS Neurosci Ther 2011; 17: 742-749.
- 18. Winhoven S, Coulson I, Bottomley W. Brachioradial pruritus: response to treatment with gabapentin. Br J Dermatol 2004; 150: 786-787
- 19. Porzio G, Aielli F, Verna L, Porto C, Tudini M, Cannita K, et al. Efficacy of pregabalin in the management of cetuximabrelated itch. J Pain Symptom Manage 2006; 32: 397-398.
- 20. Yesudian P, Wilson N. Efficacy of gabapentin in the management of pruritus of unknown origin. Arch Dermatol 2005; 141: 1507-1509.
- 21. Yosipovitch G, Samuel LS. Neuropathic and psychogenic itch. Dermatol Ther 2008; 21: 32-41.
- 22. Sakurada T, Manome Y, Tan-No K, Sakurada S, Kisara K. The effects of substance P analogues on the scratching, biting and licking response induced by intrathecal injection of N-methyld-aspartate in mice. Br J Pharmacol 1990; 101: 307-310.
- 23. Horvath G, Joo G, Kekesi G, Farkas I, Tuboly G, Petrovszki Z, et al. Inhibition of itch-related responses at spinal level in rats. Acta Physiol Hung 2011; 98: 480-490.
- 24. Jinks SL, Carstens E, Spinal NMDA receptor involvement in expansion of dorsal horn neuronal receptive field area produced by intracutaneous histamine. J Neurophysiol 1998; 79: 1613-1618.
- 25. Andoh T, Kuraishi Y. Nitric oxide enhances substance Pinduced itch-associated responses in mice. Br J Pharmacol 2003; 138: 202-208.
- 26. Ostadhadi S, Haj-Mirzaian A, Azimi E, Mansouri P, Dehpour A. Involvement of nitric oxide in serotonin-induced scratching in mice. Clin Exp Dermatol 2015; 40: 647-652.
- 27. Srebro DP, Vučković SM, Vujović KRS, Prostran MŠ. TRPA1, NMDA receptors and nitric oxide mediate mechanical hyperalgesia induced by local injection of magnesium sulfate into the rat hind paw. Physiol Behav 2015; 139: 267-273.
- 28. Freire MAM, Guimarães JS, Leal WG, Pereira A. Pain modulation by nitric oxide in the spinal cord. Front Neurosci 2009; 3: 175.
- 29. Meller S, Gebhart G. Nitric oxide (NO) and nociceptive proces-

www.medicaljournals.se/acta

sing in the spinal cord. Pain 1993; 52: 127-136.

- 30. Foroutan A, Haddadi NS, Ostadhadi S, Sistany N, Dehpour AR. Chloroquine-induced scratching is mediated by NO/ cGMP pathway in mice. Pharmacol Biochem Behav 2015; 134: 79-84.
- 31. Fawcett A. Guidelines for the Housing of Mice in Scientific Institutions. In: NSW Department of Primary Industries AWUWPH, ed.: New South Wales Government, 2012.
- 32. Vikelis M, Mitsikostas DD. The role of glutamate and its receptors in migraine, CNS Neurol Disord Drug Targets 2007; 6: 251-257.
- 33. Ghasemi M, Raza M, Dehpour AR, NMDA receptor antagonists augment antidepressant-like effects of lithium in the mouse forced swimming test. J Psychopharmacol 2010; 24: 585-594
- 34. Miller KE, Hoffman EM, Sutharshan M, Schechter R. Glutamate pharmacology and metabolism in peripheral primary afferents: physiological and pathophysiological mechanisms. Pharmacol Ther 2011; 130: 283-309.
- 35. Kaster MP, Ferreira PK, Santos AR, Rodrigues AL. Effects of potassium channel inhibitors in the forced swimming test: possible involvement of L-arginine-nitric oxide-soluble guanylate cyclase pathway. Behav Brain Res 2005; 165: 204-209.
- 36. Bryan NS, Grisham MB. Methods to detect nitric oxide and its metabolites in biological samples. Free Radic Biol Med 2007: 43: 645-657.
- 37. Schlosburg JE, Boger DL, Cravatt BF, Lichtman AH. Endocannabinoid modulation of scratching response in an acute allergenic model: a new prospective neural therapeutic target for pruritus. J Pharmacol Exp Ther 2009; 329: 314-323.
- 38. Tsukumo Y, Andoh T, Yamaguchi T, Nojima H, Kuraishi Y. Involvement of nitric oxide in itch-scratch response of NC mice. Nihon Yakurigaku Zasshi. Folia Pharmacol Jpn 1999; 114: 17-21.
- 39. Han L. Dong X. Itch mechanisms and circuits. Annu Rev Biophys 2014; 43: 331.
- McNamara CR, Mandel-Brehm J, Bautista DM, Siemens J, 40. Deranian KL, Zhao M, et al. TRPA1 mediates formalin-induced pain. Proc Natl Acad Sci U S A 2007; 104: 13525-13530.
- 41. Bleakman D, Alt A, Nisenbaum ES. Glutamate receptors and pain. Seminars in cell & developmental biology; 2006: Elsevier; 2006. p. 592-604.
- 42. Omote K, Kawamata T, Kawamata M, Namiki A. Formalininduced release of excitatory amino acids in the skin of the rat hindpaw. Brain Res 1998; 787: 161-164.
- 43. Akiyama T, Tominaga M, Takamori K, Carstens MI, Carstens E. Roles of glutamate, substance P, and gastrin-releasing peptide as spinal neurotransmitters of histaminergic and nonhistaminergic itch. Pain 2014; 155: 80-92.
- 44. Traynelis SF, Wollmuth LP, McBain CJ, Menniti FS, Vance KM, Ogden KK, et al. Glutamate receptor ion channels: structure, regulation, and function. Pharmacol Rev 2010; 62: 405-496.
- 45. Malmberg AB, Yaksh TL. Spinal nitric oxide synthesis inhibition blocks NMDA-induced thermal hyperalgesia and produces antinociception in the formalin test in rats. Pain 1993; 54: 291-300.
- 46. Dzoljic E, De Vries R, Dzoljic M. New and potent inhibitors of nitric oxide synthase reduce motor activity in mice. Behav Brain Res 1997; 87: 209-212.
- 47. Harkin A, Connor TJ, Burns MP, Kelly JP. Nitric oxide synthase inhibitors augment the effects of serotonin re-uptake inhibitors in the forced swimming test. Eur Neuropsychopharmacol 2004; 14: 274-281.
- 48. Hölscher C. Nitric oxide, the enigmatic neuronal messenger: its role in synaptic plasticity. Trends Neurosci 1997; 20: 298-303.
- 49. Nelson RJ, Demas GE, Huang PL, Fishman MC, Dawson VL, Dawson TM, et al. Behavioural abnormalities in male mice lacking neuronal nitric oxide synthase. Nature 1995; 378: 383-386.
- 50. Wiley JL, Cristello AF, Balster RL. Effects of site-selective NMDA receptor antagonists in an elevated plus-maze model of anxiety in mice. Eur J Pharmacol 1995; 294: 101-107.
- 51. Urbonas A, Schwartz RA, Szepietowski JC. Uremic pruritus:

ActaDV

an update. Am J Nephrol 2001; 21: 343-350.

- 52. Ormerod A, Weller R, Copeland P, Benjamin N, Ralston S, Grabowksi P, et al. Detection of nitric oxide and nitric oxide synthases in psoriasis. Arch Dermatol Res 1998; 290: 3-8.
- 53. Taniuchi S, Kojima T, Hara Mt K, Yamamoto A, Sasai M, Takahashi H, et al. Increased serum nitrate levels in infants with atopic dermatitis. Allergy 2001; 56: 693-695.
- 54. Meller ST, Lewis SJ, Bates JN, Brody MJ, Gebhart G. Is there a role for an endothelium-derived relaxing factor in nociception? Brain Res 1990; 531: 342-345.
- 55. Takahashi N, Mizuno Y, Kozai D, Yamamoto S, Kiyonaka S, Shibata T, et al. Molecular characterization of TRPA1 channel activation by cysteine-reactive inflammatory mediators. Channels 2008; 2: 287-298.
- 56. Yoshida T, Inoue R, Morii T, Takahashi N, Yamamoto S, Hara Y, et al. Nitric oxide activates TRP channels by cysteine Snitrosvlation. Nat Chem Biol 2006; 2: 596-607.
- 57. Purcell W, Doyle K, Westgate C, Atterwill C. Characterisation of a functional polyamine site on rat mast cells: association with a NMDA receptor macrocomplex. J Neuroimmunol 1996; 65: 49-53.
- 58. Coggeshall RE, Carlton SM. Ultrastructural analysis of NMDA, AMPA, and kainate receptors on unmyelinated and myelina-

- ted axons in the periphery. J Comp Neurol 1998; 391: 78-86. 59. Sasaki M, Yamaoka J, Mivachi Y. The effect of ultraviolet B irradiation on nitric oxide synthase expression in murine keratinocytes. Exp Dermatol 2000; 9: 417-422.
- 60. Shimizu Y, Sakai M, Umemura Y, Ueda H. Immunohistochemical Localization of nitric oxide synthase in normal human skin: expression of endothelial-type and inducible-type nitric oxide synthase in keratinocytes. J Dermatol 1997; 24: 80-87.
- 61. Thippeswamy T, Jain R, Mumtaz N, Morris R. Inhibition of neuronal nitric oxide synthase results in neurodegenerative changes in the axotomised dorsal root ganglion neurons: evidence for a neuroprotective role of nitric oxide in vivo. Neurosci Res 2001; 40: 37-44.
- 62. Kwan KY, Glazer JM, Corey DP, Rice FL, Stucky CL. TRPA1 modulates mechanotransduction in cutaneous sensory neurons. J Neurosci 2009; 29: 4808-4819.
- 63. Oh M-H, Oh SY, Lu J, Lou H, Myers AC, Zhu Z, et al. TRPA1dependent pruritus in IL-13-induced chronic atopic dermatitis. J Immunol 2013; 191: 5371-5382.
- 64. Green K, Lim H. Effects of chloroquine on release of mediators from mast cells. Skin Pharmacol Physiol 1989; 2: 77-85.
- 65. Nosal R. Drabikova K. Pecivova J. Effect of chloroquine on isolated mast cells. Agents Actions 1991; 33: 37-40.